

脂肪干细胞外泌体功能研究进展

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摘要: 脂肪干细胞外泌体(ADSCs-Exos)不含活细胞, 抗原性低, 性质稳定, 在血管生成、组织再生、免疫调节等方面发挥了重要作用。作为一种无细胞疗法, ADSCs-Exos在组织修复与再生等方面具有广阔的应用前景。该文就ADSCs-Exos功能研究进展作一综述。

关键词: 脂肪干细胞; 外泌体; miRNA

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Progress on function of adipose-derived stem cell exosomes

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Abstract: Adipose-derived stem cell exosomes (ADSC-Exos) are cell-free and stable with low antigenicity, which plays an important role in angiogenesis, tissue regeneration, immune regulation, and so on. As a cell-free therapy, ADSCs-Exos have broad prospects in tissue repair and regeneration. This article reviewed the progress on functional of ADSCs-Exos.

Key words: adipose-derived stem cell; exosome; miRNA

脂肪干细胞(ADSCs)具有分化成不同细胞系的能力, 还具有促进内皮细胞、纤维细胞迁移和增殖等功能^[1]。研究表明, ADSCs主要通过其外泌体发挥作用^[2]。外泌体是一种直径小于150 nm的细胞外囊泡结构, 将蛋白质或核酸运输到需要修复的靶细胞, 介导局部和系统的细胞间通信和信号转导, 调节受损组织细胞的多种生物学行为^[3]。相比于ADSCs, 脂肪干细胞外泌体(ADSCs-Exos)具有性质更稳定、便于管理和运输、不含活细胞、生物学功能不随时间延长而

衰减、低抗原性等优点^[1,4]。因此ADSCs-Exos作为一种无细胞疗法, 在组织修复与再生等方面具有广阔的应用前景。本文就ADSCs-Exos的研究成果及现状作一综述。

1 促进伤口愈合和血管新生

1.1 促进成纤维细胞增殖、迁移及分化

Hu^[5]和Zhang^[6]等发现ADSCs-Exos通过增加N-钙粘蛋白、cyclin-1、增殖细胞核抗原和I、III型胶原蛋白基因的表达, 或通过PI3K/Akt信号通路促进胶原的合成, 刺激成纤维细胞增殖与迁移, 促进皮肤无瘢痕修复。Ren等^[7]研究报道ADSCs-Exos可显著上调成纤维细胞增殖标志物(cyclin D1、cyclin D2、cyclin A1、cyclin A2)的表达, 能够显著促进成纤维细胞以及过氧化氢诱导的表皮角质细胞、人皮肤成纤维细胞的增殖与迁移。进一步研究发现透明质酸结合ADSCs-Exos能够明显促进急性皮肤伤口的愈合^[8]。

1.2 促进血管内皮细胞增殖、迁移和分化

研究表明ADSCs-Exos可上调生长因子(VEGF-A、PDGFA、EGF、FGF2)的表达, 在体内外显著促进人脐静脉血管内皮细胞的增殖、迁移及血管形成^[1,7], 且低氧情况下效果更加显著^[9]。同样在缺氧^[10]或缺糖

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损伤情况下^[11],ADSCs-Exos的miR-181b、miR-126可促进内皮祖细胞的迁移和血管生成,且在过氧化氢存在的情况下更有利于血管的生成^[12]。ADSCs-Exos富含的miR-125a和miR-31,可转移到血管内皮细胞中刺激细胞增殖以及促进血管生成^[13]。此外,ADSC-Exos还能促进皮瓣的血管化^[14]。ADSCs-Exos促进血管新生的作用为缺血再灌注损伤、人工皮肤中皮瓣新生血管不足、愈合不良的治疗提供了新策略。

1.3 调节胶原合成,减少疤痕形成

Yang等^[15]发现,ADSCs-Exos高表达的miRNA-21可以增强胶原合成,优化胶原沉积,显著改善小鼠全层皮肤伤口的愈合效果。ADSCs-Exos早期通过I、III型胶原的合成促进胶原重塑,晚期则通过抑制胶原形成减少疤痕形成^[5]。Wang等^[16]发现具有ADSCs-Exos的水凝胶和水凝胶支架载体材料释放的ADSCs-Exos可以显著增加皮肤附件的再生,减少疤痕组织的形成。

2 协调成骨与破骨作用

ADSCs-Exos具有显著的成骨诱导能力^[17]。体外研究证实ADSCs-Exos可通过减少核因子 κ B配体受体激活剂的表达从而拮抗缺血、缺氧引起的骨细胞凋亡,减少破骨细胞生成^[18]。研究表明多巴胺的羟基乙酸共聚物(PLGA)/聚多巴胺(PDA)^[19]、 β -磷酸三钙支架^[20]与ADSC-Exos组合,可增强ADSC-Exos促进人骨髓来源干细胞成骨、增殖和迁移的能力。此外,ADSCs-Exos高表达的miR-375可以促进骨再生,且50 mg/L的Exos(miR-375)可促进人骨髓间充质干细胞的成骨分化^[21]。

3 双向调节作用

3.1 细胞凋亡中的抑制与激活

ADSCs-Exos在心脏、神经系统、骨组织、肾脏等损伤以及血清饥饿处理或化疗药物5-氟尿嘧啶诱导的乳腺癌中具有抑制细胞凋亡的作用,而其在转移性前列腺癌、A2780和SKOV-3卵巢癌细胞中则起到激活细胞凋亡的作用。ADSCs-Exos可抑制缺血再灌注损伤^[12]、氧化应激^[22]、缺氧^[23]情况下的心肌细胞凋亡。在缺血再灌注损伤情况下,Bai等^[12]研究中指出,在过氧化氢刺激下可促进ADSCs-Exos抑制心肌细胞凋亡的作用。在小鼠肾脏缺血再灌注损伤中,ADSCs-Exos抑制肾脏细胞凋亡,促进肾脏细胞增殖^[24]。ADSCs-Exos可通过上调Bcl-2/Bax抑制坐骨神经损伤部位的施旺细胞(SCs)^[25]、血清剥夺诱导的骨样细胞系

MLO-Y4细胞^[18]的凋亡。在各类癌细胞中,Wang等^[26]指出ADSCs-Exos可激活Hippo信号通路的两个关键下游效应蛋白YAP和TAZ,在体外促进乳腺癌细胞的增殖和迁移,并保护其免受血清饥饿处理或化疗药物5-氟尿嘧啶诱导的早、晚期癌细胞的凋亡。但ADSCs-Exos对不同类型癌细胞的效果不同。如ADSCs-Exos分泌的miR-145可通过caspase-3/7途径抑制转移性前列腺癌细胞增殖,诱导癌细胞凋亡^[27]。Reza等^[28]则证明了ADSCs-Exos通过诱导细胞周期S、G2期阻滞,从而抑制A2780和SKOV-3卵巢癌细胞的增殖与集落形成能力,促进其凋亡。

3.2 细胞自噬中的抑制与激活

ADSCs-Exos在细胞自噬中具有双向作用,如在肝受损细胞和糖尿病、肾病的足细胞中起到激活自噬的作用,而在坐骨神经损伤的SCs中起到抑制自噬的作用。Qu等^[29]指出ADSCs-Exos能选择性转移miR-181-5p到受损的肝细胞,通过抑制其STAT3/Bcl-2/Beclin 1途径增加自噬,有效治疗肝病。也有研究表明ADSCs-Exos携带的miR-486可以抑制足细胞中Smad1/mTOR信号通路的激活,导致自噬增加以及足细胞凋亡减少^[30]。相反,ADSCs-Exos通过减轻坐骨神经损伤部位的SCs自噬作用,还可以加速周围神经损伤的修复^[31]。

4 免疫反应中的作用

4.1 抗炎

ADSCs-Exos可介导miR-21-3p、miR-17、miR-223等抑制炎症反应。其中ADSCs-Exos通过miR-21-3p抑制缺氧/再氧化处理细胞的凋亡,还可通过miR-21-3p上调MAT2B,进而抑制脑组织的凋亡和炎症反应^[32-33]。ADSC-Exos也可介导miR-17和miR-223发挥作用,通过靶向硫氧还原蛋白互作蛋白(TXNIP)和抑制肝巨噬细胞炎症小体激活,减少炎症的发生^[34]。另有研究报道ADSCs-Exos可通过抑制NF- κ B和MAPK信号通路,抑制BV2小胶质细胞的活化,从而抑制TNF- α 、IL-6、IL-1 β 、iNOS等炎症因子的基因表达和蛋白分泌,保护细胞免受损伤^[35-36]。在IL-1 β 刺激的关节炎软骨细胞中,ADSCs-Exos使核因子- κ B和激活蛋白1活化降低,增强抗炎细胞因子IL-10的产生和膜联蛋白A1的表达,减少炎症反应^[37]。此外ADSCs-Exos可诱导巨噬细胞向抗炎M2细胞极化^[38],并促使其表达高水平的酪氨酸羟化酶、精氨酸酶-1和IL-10^[39]。ADSCs-Exos还可以通过调节Treg细胞数量、降低血脂和炎症因子,缓解葡聚糖硫酸钠

诱导的急性结肠炎模型小鼠的症状^[40]。

4.2 免疫调节

ADSCs-Exos具有免疫调节作用,可降低体内的IFN- γ 、IL-1 β 、TNF等的水平,抑制体外刺激下T细胞的增殖、分化及活化,调节机体的免疫功能,促进组织修复^[41-42]。此外,ADSCs-Exos携带的miR-223可诱导巨噬细胞极化,具有免疫调节的作用,且低氧条件下作用更显著^[43]。在血管复合异体移植术中,ADSCs-Exos处理后的供体细胞嵌合性、Tr1、Treg明显增加,CD4T、Th1细胞明显减少,抑制同种异体免疫反应^[44]。

5 作为靶向治疗的转运载体

研究表明含有聚吡咯钛的ADSCs-Exos可以显著促进体内成骨^[45]。Li等^[46]以ADSCs-Exos为载体,将CD63-血管内皮生长因子靶向输送到淋巴内皮细胞,显著促进了淋巴内皮细胞的增殖、迁移和淋巴管形成,从而治疗淋巴水肿。此外Zhang等^[47]研究发现,ADSCs-Exos是miRNAs或circRNAs的重要载体,在非小细胞肺癌中,circ-100395过表达的ADSCs-Exos可以抑制非小细胞肺癌细胞的生物活性,从而抑制非小细胞肺癌的恶性转化。而载有miR-381的ADSCs-Exos可在体外抑制人乳腺癌细胞系MDA-MB-231的增殖、迁移和侵袭能力^[48]。研究表明,对奥沙利铂耐药的结直肠癌细胞Exos可将circ-0005963(即ciRS-122)传递给敏感细胞,促进糖酵解和耐药性,而Exos转运的si-ciRS-122在体内可通过调控ciRS-122-miR-122-PKM2通路抑制糖酵解,逆转对奥沙利铂的耐药^[49]。同样,ADSCs-Exos是否可以通过某些机制参与调控细胞的耐药性也值得我们探讨,这将为癌症耐药性治疗提供新思路。

6 其他作用

除上述的功能外,ADSCs-Exos在调节肥胖、调控胰岛素抵抗、促进组织再生等方面也具有一定功能。ADSCs-Exos可以转移到巨噬细胞中诱导抗炎M2表型,增加解偶联蛋白1的表达,促进白色脂肪组织的形成,从而改善肥胖相关的炎症和代谢。在肥胖小鼠模型中,ADSCs-Exos可以降低肥胖引起的全身胰岛素抵抗和肝脏脂肪变性^[39]。ADSCs-Exos可以通过刺激PI3K-Akt、Jak-STAT和Wnt等与神经再生相关的信号通路,促进SCs的增殖、迁移、分化,进而促进神经再生^[50-51]。ADSCs-Exos还可以通过激活内皮细胞中PI3K/AKT信号通路减轻组蛋白诱导的内皮损伤,从而改善急性肺损伤^[52]。此外,Wu等^[53]研究发现

ADSCs-Exos可以促进毛囊的发育与再生,促进头发的再生。

7 小结与展望

本文通过总结ADSCs-Exos的功能和研究现状,发现ADSCs-Exos在血管重建、组织修复与再生、抑制肿瘤生长、靶向治疗等领域有较大的发展空间,其活性和生物学功能受环境影响。如在低氧环境或过氧化氢刺激下,ADSCs-Exos促血管新生和免疫调节的效果更加显著。此外,ADSCs-Exos可作为靶向治疗的转运载体,利于定向治疗,减少副作用。其联合治疗效果较为明显,如透明质酸、多巴胺的PLGA、 β -磷酸三钙和水凝胶支架等与ADSCs-Exos结合,可以更好地发挥作用,这些都为ADSCs-Exos的功能优化提供了策略。

ADSCs-Exos具有双向调节作用,如促进或抑制细胞凋亡及自噬、对炎症反应和免疫调节的正负调控等。在损伤情况下,为实现组织修复再生,ADSCs-Exos往往起到抑制细胞凋亡的作用;但在癌细胞中,ADSCs-Exos既可抑制细胞凋亡,也可促进凋亡,这一作用可能取决于癌症的类型或ADSCs-Exos中miRNAs的种类。如前所述,ADSCs-Exos抑制乳腺癌细胞凋亡时,伴随有Hippo信号通路的两个关键下游效应蛋白YAP和TAZ的激活,而miR-151等家族成员被证实具有促癌功能^[26]。那么ADSCs-Exos是否通过介导miR-151抑制乳腺癌细胞凋亡,ADSCs-Exos与miR-151之间与Hippo信号通路效应蛋白YAP和TAZ是否存在联系,如何利用好ADSCs-Exos这把“双刃剑”调节肿瘤生长,还有待进一步研究。

参考文献:

- [1]张静,易阳艳,阳水发,等. 脂肪干细胞来源外泌体对人脐静脉血管内皮细胞增殖、迁移及管样分化的影响[J]. 中国修复重建外科杂志, 2018, 32(10):1351-1357.
- [2]KHAN M, KISHORE R. Stem cell exosomes: Cell-free therapy for organ repair[J]. *Methods Mol Biol*, 2017, 1553:315-321.
- [3]HONG P, YANG H, WU Y, et al. The functions and clinical application potential of exosomes derived from adipose mesenchymal stem cells: A comprehensive review[J]. *Stem Cell Res Ther*, 2019, 10(1):242.
- [4]XIONG M, ZHANG Q, HU W, et al. Exosomes from adipose-derived stem cells: The emerging roles and applications in tissue regeneration of plastic and cosmetic surgery[J]. *Front Cell Dev Biol*, 2020, 8:574223.
- [5]HU L, WANG J, ZHOU X, et al. Exosomes derived from

- human adipose mesenchymal stem cells accelerates cutaneous wound healing via optimizing the characteristics of fibroblasts[J]. *Sci Rep*, 2016,6(1):32993.
- [6] ZHANG W, BAI X, ZHAO B, et al. Cell-free therapy based on adipose tissue stem cell-derived exosomes promotes wound healing via the PI3K/Akt signaling pathway[J]. *Exp Cell Res*, 2018, 370(2):333-342.
- [7] REN S, CHEN J, DUSCHER D, et al. Microvesicles from human adipose stem cells promote wound healing by optimizing cellular functions via AKT and ERK signaling pathways[J]. *Stem Cell Res Ther*, 2019, 10(1):47.
- [8] LIU K, CHEN C, ZHANG H, et al. Adipose stem cell-derived exosomes in combination with hyaluronic acid accelerate wound healing through enhancing re-epithelialization and vascularization[J]. *Br J Dermatol*, 2019, 181(4):854-856.
- [9] HAN Y, REN J, BAI Y, et al. Exosomes from hypoxia-treated human adipose-derived mesenchymal stem cells enhance angiogenesis through VEGF/VEGF-R[J]. *Int J Biochem Cell B*, 2019, 109:59-68.
- [10] LUO Q, GUO D, LIU G, et al. Exosomes from miR-126-overexpressing adscs are therapeutic in relieving acute myocardial ischaemic injury[J]. *Cell Physiol Biochem*, 2018, 44(6):2105-2116.
- [11] YANG Y, CAI Y, ZHANG Y, et al. Exosomes secreted by adipose-derived stem cells contribute to angiogenesis of brain microvascular endothelial cells following oxygen-glucose deprivation in vitro through microRNA-181b/TRPM7 axis[J]. *J Mol Neurosci*, 2018, 65(1):74-83.
- [12] BAI Y, HAN Y, YAN X, et al. Adipose mesenchymal stem cell-derived exosomes stimulated by hydrogen peroxide enhanced skin flap recovery in ischemia-reperfusion injury[J]. *Biochem Biophys Res Commun*, 2018, 500(2): 310-317.
- [13] LIANG X, ZHANG L, WANG S, et al. Exosomes secreted by mesenchymal stem cells promote endothelial cell angiogenesis by transferring miR-125a[J]. *J Cell Sci*, 2016, 129(11):2182-2189.
- [14] XIONG J, LIU Z, WU M, et al. Comparison of proangiogenic effects of adipose-derived stem cells and foreskin fibroblast exosomes on artificial dermis prefabricated flaps[J]. *Stem Cells Int*, 2020, 2020:1-14.
- [15] YANG C, LUO L, BAI X, et al. Highly-expressed microRNA-21 in adipose derived stem cell exosomes can enhance the migration and proliferation of the HaCaT cells by increasing the MMP-9 expression through the PI3K/AKT pathway[J]. *Arch Biochem Biophys*, 2020, 681:108259.
- [16] WANG C, WANG M, XU T, et al. Engineering bioactive self-healing antibacterial exosomes hydrogel for promoting chronic diabetic wound healing and complete skin regeneration[J]. *Theranostics*, 2019, 9(1):65-76.
- [17] BORCIANI G, MONTALBANO G, BALDINI N, et al. Co-culture systems of osteoblasts and osteoclasts: Simulating in vitro bone remodeling in regenerative approaches[J]. *Acta Biomater*, 2020, 108:22-45.
- [18] REN L, SONG Z, CAI Q, et al. Adipose mesenchymal stem cell-derived exosomes ameliorate hypoxia/serum deprivation-induced osteocyte apoptosis and osteocyte-mediated osteoclastogenesis in vitro[J]. *Biochem Biophys Res Commun*, 2019, 508(1):138-144.
- [19] LI W, LIU Y, ZHANG P, et al. Tissue-engineered bone immobilized with human adipose stem cells-derived exosomes promotes bone regeneration[J]. *ACS Appl Mater Inter*, 2018, 10(6):5240-5254.
- [20] ZHANG J, LIU X, LI H, et al. Exosomes/tricalcium phosphate combination scaffolds can enhance bone regeneration by activating the PI3K/Akt signaling pathway[J]. *Stem Cell Res Ther*, 2016, 7(1):136.
- [21] CHEN S, TANG Y, LIU Y, et al. Exosomes derived from miR-375-overexpressing human adipose mesenchymal stem cells promote bone regeneration[J]. *Cell Proliferat*, 2019, 52(5): e12669.
- [22] LIU Z, XU Y, WAN Y, et al. Exosomes from adipose-derived mesenchymal stem cells prevent cardiomyocyte apoptosis induced by oxidative stress[J]. *Cell Death Discov*, 2019, 5(1):79.
- [23] XU H, WANG Z, LIU L, et al. Exosomes derived from adipose tissue, bone marrow, and umbilical cord blood for cardioprotection after myocardial infarction[J]. *J Cell Biochem*, 2020, 121(3):2089-2102.
- [24] LIN K C, YIP H K, SHAO P L, et al. Combination of adipose-derived mesenchymal stem cells (ADMSC) and ADMSC-derived exosomes for protecting kidney from acute ischemia-reperfusion injury[J]. *Int J Cardiol*, 2016, 216:173-185.
- [25] LIU C Y, YIN G, SUN Y D, et al. Effect of exosomes from adipose-derived stem cells on the apoptosis of Schwann cells in peripheral nerve injury[J]. *CNS Neurosci Ther*, 2020, 26(2):189-196.
- [26] WANG S, SU X, XU M, et al. Exosomes secreted by mesenchymal stromal/stem cell-derived adipocytes promote breast cancer cell growth via activation of Hippo signaling pathway[J]. *Stem Cell Res Ther*, 2019, 10(1):117.
- [27] TAKAHARA K, II M, INAMOTO T, et al. MicroRNA-145 mediates the inhibitory effect of adipose tissue-derived stromal cells on prostate cancer[J]. *Stem Cells Dev*, 2016, 25(17):1290-1298.
- [28] REZA A M M T, CHOI Y, YASUDA H, et al. Human adipose mesenchymal stem cell-derived exosomal-miRNAs are critical factors for inducing anti-proliferation signalling to A2780 and SKOV-3 ovarian cancer cells[J]. *Sci Rep*, 2016, 6(1): 38498.
- [29] QU Y, ZHANG Q, CAI X, et al. Exosomes derived from miR-181-5p-modified adipose-derived mesenchymal stem cells prevent liver fibrosis via autophagy activation[J]. *J Cell Mol Med*, 2017, 21(10):2491-2502.
- [30] JIN J, SHI Y, GONG J, et al. Exosome secreted from adipose-

- derived stem cells attenuates diabetic nephropathy by promoting autophagy flux and inhibiting apoptosis in podocyte[J]. *Stem Cell Res Ther*, 2019, 10(1):95.
- [31] YIN G, YU B, LIU C, et al. Exosomes produced by adipose-derived stem cells inhibit schwann cells autophagy and promote the regeneration of the myelin sheath[J]. *Int J Biochem Cell Biol*, 2021, 132:105921.
- [32] LI C, FEI K, TIAN F, et al. Adipose-derived mesenchymal stem cells attenuate ischemic brain injuries in rats by modulating miR-21-3p/MAT2B signaling transduction[J]. *Croat Med J*, 2019, 60(5):439-448.
- [33] GE X, LI W, HUANG S, et al. Increased miR-21-3p in injured brain microvascular endothelial cells after traumatic brain injury aggravates blood-brain barrier damage by promoting cellular apoptosis and inflammation through targeting MAT2B[J]. *J Neurotrauma*, 2019, 36(8):1291-1305.
- [34] LIU Y, LOU G, LI A, et al. AMSC-derived exosomes alleviate lipopolysaccharide/d-galactosamine-induced acute liver failure by miR-17-mediated reduction of TXNIP/NLRP3 inflammasome activation in macrophages[J]. *EBioMedicine*, 2018, 36:140-150.
- [35] FENG N, JIA Y, HUANG X. Exosomes from adipose-derived stem cells alleviate neural injury caused by microglia activation via suppressing NF- κ B and MAPK pathway[J]. *J Neuroimmunol*, 2019, 334:576996.
- [36] SONG Y, LI H, REN X, et al. SNHG9, delivered by adipocyte-derived exosomes, alleviates inflammation and apoptosis of endothelial cells through suppressing TRADD expression[J]. *Eur J Pharmacol*, 2020, 872:172977.
- [37] TOFINO-VIAN M, GUILLEN M I, PEREZ DEL CAZ M D, et al. Microvesicles from human adipose tissue-derived mesenchymal stem cells as a new protective strategy in osteoarthritic chondrocytes[J]. *Cell Physiol Biochem*, 2018, 47(1): 11-25.
- [38] SHEN K, JIA Y, WANG X, et al. Exosomes from adipose-derived stem cells alleviate the inflammation and oxidative stress via regulating Nrf2/HO-1 axis in macrophages[J]. *Free Radic Biol Med*, 2021, 165:54-66.
- [39] ZHAO H, SHANG Q, PAN Z, et al. Exosomes from adipose-derived stem cells attenuate adipose inflammation and obesity through polarizing M2 macrophages and beiging in white adipose tissue[J]. *Diabetes*, 2018, 67(2):235-247.
- [40] HEIDARI N, ABBASI-KENARSARI H, NAMAKI S, et al. Adipose-derived mesenchymal stem cell-secreted exosome alleviates dextran sulfate sodium-induced acute colitis by Treg cell induction and inflammatory cytokine reduction[J]. *J Cell Physiol*, 2021, 236(8):5906-5920.
- [41] KORDELAS L, REBMANN V, LUDWIG A K, et al. MSC-derived exosomes: A novel tool to treat therapy-refractory graft-versus-host disease[J]. *Leukemia*, 2014, 28(4):970-973.
- [42] BLAZQUEZ R, SANCHEZ-MARGALLO F M, DE LA ROSA O, et al. Immunomodulatory potential of human adipose mesenchymal stem cells derived exosomes on in vitro stimulated T cells[J]. *Front Immunol*, 2014, 5:556.
- [43] LO S C, REVERBERI D, BALBI C, et al. Mesenchymal stem cell-derived extracellular vesicles as mediators of anti-inflammatory effects: Endorsement of macrophage polarization[J]. *Stem Cells Transl Med*, 2017, 6(3):1018-1028.
- [44] CHEN Z, XUE S, ZHANG S, et al. Exosomes from donor-derived adipose mesenchymal stem cells prolong the survival of vascularized composite allografts[J]. *J Cell Physiol*, 2021, 236(8):5895-5905.
- [45] CHEN L, MOU S, LI F, et al. Self-assembled human adipose-derived stem cell-derived extracellular vesicle-functionalized biotin-doped polypyrrole titanium with long-term stability and potential osteoinductive ability[J]. *ACS Appl Mater Interfaces*, 2019, 11(49):46183-46196.
- [46] LI B, YANG J, WANG R, et al. Delivery of vascular endothelial growth factor (VEGFC) via engineered exosomes improves lymphedema[J]. *Ann Transl Med*, 2020, 8(22):1498.
- [47] ZHANG C, CAO J, LV W, et al. CircRNA_100395 carried by exosomes from adipose-derived mesenchymal stem cells inhibits the malignant transformation of non-small cell lung carcinoma through the miR-141-3p-LATS2 axis[J]. *Front Cell Dev Biol*, 2021, 9:663147.
- [48] SHOJAEI S, HASHEMI S M, GHANBARIAN H, et al. Delivery of miR-381-3p mimic by mesenchymal stem cell-derived exosomes inhibits triple negative breast cancer aggressiveness; An in vitro study[J]. *Stem Cell Rev Rep*, 2021, 17(3):1027-1038.
- [49] WANG X, ZHANG H, YANG H, et al. Exosome-delivered circRNA promotes glycolysis to induce chemoresistance through the miR-122-PKM2 axis in colorectal cancer[J]. *Mol Oncol*, 2020, 14(3):539-555.
- [50] NI J, LI H, ZHOU Y, et al. Therapeutic potential of human adipose-derived stem cell exosomes in stress urinary incontinence-an in vitro and in vivo study[J]. *Cell Physiol Biochem*, 2018, 48(4):1710-1722.
- [51] CHEN J, REN S, DUSCHER D, et al. Exosomes from human adipose-derived stem cells promote sciatic nerve regeneration via optimizing Schwann cell function[J]. *J Cell Physiol*, 2019, 234(12):23097-23110.
- [52] MIZUTA Y, AKAHOSHI T, GUO J, et al. Exosomes from adipose tissue-derived mesenchymal stem cells ameliorate histone-induced acute lung injury by activating the PI3K/Akt pathway in endothelial cells[J]. *Stem Cell Res Ther*, 2020, 11(1):508.
- [53] WU J, YANG Q, WU S, et al. Adipose-derived stem cell exosomes promoted hair regeneration[J]. *Tissue Eng Regen Med*, 2021, 18(4):685-691.